5'-Nucleotidase Assay Kit (5'-NT)

Method: Colorimetric (Kinetic)

Cat .No.	Size	Instrument
GB8010G	R1: 2 × 20 ml R2: 1 × 20 ml	For Hitachi 717 & ShimadzuCL7200/8000
GS8011G	R1: 2 × 20 ml R2: 1 × 20 ml	For Hitachi917 & OlympusAU640/400/600
GX8011G	R1: 2 × 20 ml R2: 1 × 20 ml	For SYNCHRON CX4-5-7-9/LX20

INTENDED USE

For the *in vitro* quantitative determination of 5'-nucleotidase activity in human serum or plasma.

CLINICAL SIGNIFICANCE

5'-NT is an enzyme catalyzing the hydrolysis of nucleoside-5'-monophosphates to nucleosides and inorganic phosphate. The enzyme is widely distributed in human and animal tissues. The activity present in sera is released from the membrane of liver cells by bile salts and has been used as a marker for liver disease^[1]. Increased enzyme levels in sera are associated with certain forms of liver disease, such as intra or extra-hepatic obstruction and particularly in cases of hepatic carcinoma as well as in mastectomy patients with recurrent metastases. The diagnostic value of 5'-NT has been shown to be superior to other liver enzymes, especially in cases of liver metastasis.

ASSAY PRINCIPLE

The 5'-NT assay is based on the enzymatic hydrolysis of 5'-monophosphate (5'-IMP) to form inosine which is converted to hypoxanthine by purine nucleoside phosphorylase (PNP). Hypoxanthine is then converted to uric acid and hydrogen peroxide (H_2O_2) by xanthine oxidase (XOD). H_2O_2 is further reacted with N-Ethyl-N-(2-hydroxy-3-sulfopropyl)-3-methylaniline (EHSPT) and 4-aminoantipyrine (4-AA) in the presence of peroxidase (POD) to generate quinone dye which is monitored in a kinetic manner. The entire enzymatic reaction scheme is shown below.

IMP +
$$H_2O$$
 $\xrightarrow{5'-NT}$ Inosine + Pi

Inosine + Pi \xrightarrow{PNP} Hypoxanthine + Ribose 1-phosphate

Hypoxanthine + $2H_2O$ + $2O_2$ \xrightarrow{XOD} Uric acid + $2H_2O_2$

$$2H_2O_2+4$$
-AA+EHSPT \longrightarrow $4H_2O+Quinone dye (λ_{max} =556 nm)$

SAMPLE COLLECTION AND PREPARATION

Serum or plasma samples.

Use fresh and non-hemolyzed serum or EDTA treated plasma samples. Serum or plasma samples are stable for a week at 4°C.

REAGENT COMPOSITION

Contents	Concentration of Solutions			
Reagent 1 (R1)				
Goods buffer (PH7.6)	100 mmol/L			
4-AA	2 mmol/L			
PNP	0.1 U/L			
XOD	0.2 U/L			
POD	0.6 U/L			
Stabilizer				
Reagent 2 (R2)				
Goods buffer (PH7.6)	100 mmol/L			
IMP	10 mmol/L			
EHSPT	2 mmol/L			

STABILITY AND PREPARATION OF REAGENTS

All reagents are ready to use.

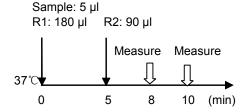
Stable up to the expiry date when stored at $2-8^{\circ}$ C. The assay kit reagents are stable for 1 month after opening and kept at $2-8^{\circ}$ C.

ASSAY PROCEDURE

Test Procedure for Analyzers (HITACHI 7170/917)

Assay Mode: 2 Point Rate 28-34

Wave Length (main/sub): 546 nm/800 nm



- 1. Mix 5 μ I sample with 180 μ I R1 and incubate at 37 $^{\circ}$ C for 5 minutes.
- 2. Add 90 μ I R2 into cuvette, mix and incubate for 3 minutes at 37 $^{\circ}$ C.
- Read initial absorbance and start timer simultaneously, read again after 1 and 2 minutes.
- 4. Calculate absorbance change per minute $(\Delta A/min)$

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CALCULATION (ε =16.18)

5'-NT (U/L) =
$$\frac{\Delta A/min \times V_t}{\epsilon \times V_s \times L}$$
 = $\Delta A/min \times 3400$

QUALITY CONTROL

For quality control, use 5'-NT control as daily quality control sera and can be purchased separately. Values should fall within a specific range. If these values fall outside the range and repetition excludes error, the following steps should be taken:

- 1. Check instrument settings and light source.
- 2. Check reaction temperature.
- 3. Check expiration date of kit and contents.

REFERENCE VALUE

Serum or plasma: 0-10 U/L

It is recommended that each laboratory establish its own reference range to reflect the age, sex, diet and geographical location of the population.

LINEARITY

The method is linear up to 300 U/L. Samples above this concentration should be diluted 1+1 with 0.9% NaCl and reassayed. Multiply the result by 2.

SPECIFIC PERFORMANCE CHARACTERISTICS INTERFERENCE

Hemoglobin≤ 500 mg/dl, Intralipid ≤1000 mg/dl ALP≤1250 U/l, without interference. Bilirubin≤40 mg/dl, Ascorbic Acid≤50 mg/dl, interference.

PRECISION

The CV of the test should be ≤10%.

Intra assay precision							
N=20	Level1	Level 2					
Mean	7.5	78.8					
SD	0.11	0.95					
CV	1.5%	1.2%					
Inter assay p	Inter assay precision						
N=5	Level1	Level 2					
Mean	7.83	79.3					
SD	0.16	1.35					
CV	2.0%	1.7%					

SENSITIVITY

The minimum detectable level that can be distinguished from zero has been determined as 1.81 U/L.

CORRELATION

This method (Y) was compared with another commercially available method (X) and the following linear regression equation obtained: Y=1.0426X+1.3074, $R^2=0.9912$; 64 patient samples were analyzed .

SAFETY PRECAUTIONS AND WARNINGS

- For *in vitro* diagnostic use only. Do not pipette by mouth. Exercise the normal precautions required for handing laboratory reagents.
- Reagents contain Sodium Azide. Avoid ingestion or contact with skin or mucous membranes. In case of skin contact, flush affected area with copious amounts of water. In case of contact with eyes or if ingested, seek immediate medical attention.
- Sodium Azide reacts with lead and copper plumbing, to form potentially explosive azides. When disposing of such reagents flush with large volumes of water to prevent azide build up. Exposed metal surfaces should be cleaned with 10% sodium hydroxide.
- 4. All specimens used in this test should be considered potentially infectious. Universal Precautions, as they apply at your facility, should be used for handling and disposing of materials during and after testing.

REFERENCES

- Eroglu A. Activities of adenosine deaminase and 5' - nucleotidase in cancerous and noncancerous human cdorectal tissues[J]. Med Oneol 2000; 17(4):319 - 24.1
- Alain Bertrand and Jean Buret, A one-step determination of 5'-nucleotidase using a centrifugal analyzer. Clinica Chimica Acta,119(1982)275-284.
- 3. By Z. Ahmed and J.L.Reis, The Activation and Inhibition of 5-Nucleotidase. Clin Chem. 1998, 69 (11), 102—106.

Web: www.bsbe.com.cn Email: tech@bsbe.com.cn

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INDEX OF SYMBOLS

Manufacture

Catalogue Number

Lot number

Date of manufacture

Use by (Expiration date)

For In-Vitro Diagnostic use only

Stored at 2-8℃

Attention: See instruction for use

Authorized Representative in the

European Company

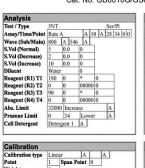


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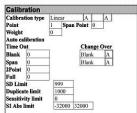
INSTRUMENT SETTINGS FOR HITACHI 917

Gcell Hitachi 7170 Parameter Application

5'-NT Cat. No: GB8010G/GS8011G/GX8011G



Range								
Applicati	ion C	od	e		*	Un	it U/L	A
Report N				5NT	5NT			
Data Mo	de				On I	Board	A	
Control I	nter	val			0			
Instrume	nt F	acti	or (Y=a)	(+b) a=	1.0	b= 0	1
Technica	Lin	nit			0		300	[
Expected	Val	ae				Qu	alitative	
	Val:	200	Α			Qu	alitative Cancel	A
	00000	Y	A	-		Qu (1)	Cancel	A
	0	Y	A	0	10	1	Cancel 0	A
(Male)	0 100	Y	A	0	10	(1)	Cancel 0 0	A
Expected (Male) (Female)	0 100	Y	A	0	10	(1)	Cancel 0 0 0	A



<standard></standard>	(1)	(2)	(3)	(4)	(5)	(6)
Concentration	0.0	0	0	0	0	0
Position	Water	0	0	0	0	0
Volume	5	0	0	0	0	0
<pre-diluent< td=""><td>-</td><td></td><td></td><td></td><td></td><td></td></pre-diluent<>	-					
Volume	0	0	0	0	0	0
Diluent	0	0	0	0	0	0
Cal. Code	0	0	0	0	0	0

K-factor = 34000

INSTRUMENT SETTINGS FOR HITACHI 902

Hitachi 7020 Instrument Settings

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Cat. No: GB8010G/GS8011G/GX8011G

No.	<chemistry></chemistry>		
1	Test Name	5'-NT	
2	Assay Code (Mthd)	Rate A	
3	Assay Code (2. Test)	0	
4	Reaction Time	10	
5	Assay Point 1	27	
6	Assay Point 2	34	
7	Assay Point 3	0	
8	Assay Point 4	0	
9	Wave Leng. (SUB)	800	
10	Wave Leng. (MAIN)	546	
11	Sample Volume	5	
12	R1 VOLUME	180	
13	R1 Pos.	*	
14	R1 Bottle Size	Large	
15	R2 VOLUME	0	
16	R2 Pos.	0	
17	R2 Bottle Size	Small	
18	R3 VOLUME	90	
19	R3 Pos.		
20	R3 Bottle Size	Small	
21	Calib. Type (Type)	K Factor	
22	Calib. Type (Wght)	0	
23	Calib. Conc. 1	0	
24	Calib. Pos. 1	99	
25	Calib. Conc. 2	0	
26	Calib. Pos. 2	0	
27	Calib. Conc. 3	0	
28	Calib. Pos. 3	0	
29	Calib. Conc. 4	0	
30	Calib. Pos. 4	0	
31	Calib. Conc. 5	0	
32	Calib. Pos. 5	0	
33	Calib. Conc. 6	0	
34	Calib. Pos. 6	0	
35	S I ABS.	0	
36	K Factor	3400	
37	K 2 Factor	10000	
38	K 3 Factor	10000	
39	K 4 Factor	10000	
40	K 5 Factor	10000	
41	A Factor	0	
42	B Factor	0	
43	C Factor	0	
44	SD Limit	999	
45	Duplicate Limit	1000	

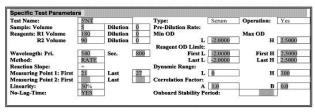
46	Sens. Limit	0	
47	S 1 ABS Limit (L)	-32000	
48	S 1 ABS Limit (H)	32000	
49	ABS Limit	32000	
50	ABS Limit (D/I)	Increase	
51	Prz. Limit	0	
52	Prz. Limit (U/D)	Lower	
53	Prz. (End Point)	35	
54	Expect. Value (L)	0	
55	Expect. Value (H)	10	
56	Instr. Fact. (a)	1	
57	Instr. Fact. (b)	0	
58	Key Setting	*	

Data entry by the user

INSTRUMENT SETTINGS FOR Olympus400/640/2700

Olympus AU640/400/2700 Instrument Settings Gcell

5'-NT Cat. No:GB8010G/GS8011G/GX8011G



Calibration Sp	ecific				
Test No.:		Name:	5'NT	Type:	SER
Cal. Type:	MB			Counts:	2
Formula:		Y = AX + B		Process:	CONC
Calibration Sel	ection: Cal. No.	OD	Conc.	Factor/OD-L	Factor/OD-H
Point 1				10000	0.000
Point 2	10000	1000			
Point 3					811111118
Point 4					
Point 5	I I I I I I I I I I I I I I I I I I I	10000	10000		
Point 6			10000	00000	
Point 7					
1-Point CalPo	int:		1000		
MB Type Facto	r;	3400			
Cali. Stability I	eriod:				

INSTRUMENT SETTINGS FOR CX4/5/7/9

Synchron CX-4/5/7/9 User-defined Chemistries

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5'-NT

Cat. No: GS8011G/GB8010G/GX8010G

USER ID:					
Chemistry Name:	5'-N7	100			
Test Name:				Calculate Factor:	6800
Reaction Type:	Rate	ī		Math Model:	Linear
Reaction Direction:				Cal Time Limit:	0 8
Units:	U/L			No. Of Calibrators:	0
Decimal Precision:	X.X				
Primary Wavelength:	560	nm		Secondary Wavelength:	700 n
Sample Volume: Primary Inject Rqt:		350000		MULTIPOINT SP	AN
A:	144	pl	#1:	1 - 2	0.000
		μl		2 - 3	0.000
Cacondary Triest Bot.			40.	3 - 4	0.000
None:	0	pl	#4:	4 - 5	0.000
Add Time:		sec	#5:	5 - 1	0.000
RAGENT BLAN				REACTION	
Start Read:				Start Read:	180 se
End Read:	300	sec		End Read:	300 se
Low ABS Limit:	-1.50	00		Low ABS Limit:	-1.500
High ABS Limit:	1.500)		High ABS Limit:	1.500
USABLE RANG	E			SUBSTRATE DEPLE	
Lower Limit:	0			Initial Rate:	
Upper Limit:	300			Delta ABS:	1.5
RECOVERY/SENSIT	IVITY				
Std Dev (conc):	*				
CV (%):	*				
Std Dev (mA):	*				
non not limit.					

Attention: * Entered By Operator

Beijing Strong Biotechnologies, Inc. Add:No. 15, North Second Street, Yanqi Economic Development Area, Huairou District, Beijing,101400,P.R.China

Tel: +86 10 6166 7168 Fax: +86 10 6166 7810 Web: www.bsbe.com.cn Email: tech@bsbe.com.cn

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